MOLECULAR DETECTION OF *inv*A AND *hil*A VIRULENT GENES IN SALMONELLA SEROVARS ISOLATED FROM FRESH WATER FISH

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Abstract: The present study was conducted to monitor the prevalence of salmonellae in fresh water fish in Gharbia governorate, Egypt. A total of 120 random samples of fresh water fish (*Tilapia niloticus*, *Mugil cephalous* and *Clarias lazera*, 40 of each) were analyzed bacteriologically for the presence of salmonella pathogens and were further identified using biochemical and serological tests. Positive samples were confirmed by polymerase chain reaction (PCR) through detection of common virulent genes invasion A (*inv*A) and hyper-invasive locus (*hil*A). The obtained biochemical and serological results revealed presence of seven different serotypes including *S. typhimurium* which was the most frequent one followed by *S. enteriditis*, *S. infantis*, *S. virchow*, *S. heidelberg*, *S. wingrove* and *S. tsevie* with a prevalence of 33.3%, 23%, 12.8%, 12.8%, 10.25%, 5% and 2.5%, respectively. The obtained molecular results revealed presence of the two virulent genes in all the detected serovars with a total prevalence rate of 100% for *inv*A gene and 66.6% for *hil*A gene. These results reflected a high prevalence of salmonella species in fresh water fish which act as a potential source of food borne infection and constitutes a major concern for public health.

Key words: fresh water fish; salmonella; serotypes; *inv*A; *hil*A; PCR

Introduction

Fish are considered as an excellent cheap source of high quality protein with very low cholesterol levels and large quantities of polyunsaturated fatty acids that cannot be synthesized by human metabolism (1, 2). However, fish are also considered as a major carrier for food borne pathogens as its natural habitat is extremely susceptible to pollution from domestic, industrial and agricultural discharges (3). Bacterial diseases in fish cause a serious threat not only to aquaculture industry but also on human health. Thus, it is not surprising to find that 12% of the food poisoning outbreaks associated with fish consumption are caused by bacteria including salmonella (4).

Salmonella, as a member of family enterobacteriacea, is a Gram negative rod shaped bacteria that contains more than 2500 serovars. Salmonella, which is considered as a common water-borne pathogen, apparently presents in the tissues of normal fish and represents no clinical signs. It can easily reach water through many...
routes, especially through sewage discharges, and can concentrate in fish tissues. It also can reach fish outside water through fish handling without following strict hygienic measures. When these contaminated fish were eaten raw or with quick inappropriate heating, the consumers would suffer from enteritis and typhoid fever. Despite extensive public health measures over the past century, salmonella remains the second leading cause of food-borne diseases worldwide (5). Accordingly, providing a safe, high quality and acceptable food to a consumer is considered a major goal for food processing industry. This can be applied by prompt and accurate isolation and characterization of salmonella and other food-borne pathogens which are the cornerstone required for proper control of food-borne diseases. This can be achieved using molecular identification of these pathogens by polymerase chain reaction (PCR) technique. The latter, is highly sensitive and more accurate method for bacterial isolation and identification, especially for salmonella sp. (6). Thus, our study was planned to identify the prevalence of salmonella species in fresh water fish in Gharbia Governorate, Egypt and to detect the common virulent genes of salmonella serovars using PCR technique.

Materials and methods

Isolation of salmonellae

A total of 120 random samples of fresh water fish including *Tilapia niloticus*, *Mugil cephalus* and *Clarias lazera* (40 of each) were collected from different markets in Gharbia governorate, Egypt. Samples were placed separately in sterile plastic bags in an ice box and transported immediately to the laboratory for bacteriological examination. According to ISO 6579 (2002) method [11]: 25 g of back muscles of each sample of fresh water fish were transferred to a homogenizer flask containing 225 ml of sterile buffered peptone (1%) and homogenized for 1 min, then incubated at 37 °C for 18 h. According to ISO 6579 (2002) method: 1 ml of each homogenate was inoculated separately into a tube of 9 ml selenite-f broth and incubated at 37°C for 18 h or 0.1 ml was transferred to 9 ml of Rappaport - Vassiliadis Soya broth (RVS) and incubated at 42°C for 24 h. A loopful from selective enriched broth was streaked onto plates of Xylose Lysine Deoxycholate (XLD) and incubated at 37 °C for 24 h. Typical pink colonies with or without black center were picked up for further identification.

Identification of salmonellae

For microscopical identification, films from suspected colonies were prepared, fixed and stained with Gram stain showing Gram negative rod-shaped, motile bacilli that indicates salmonella pathogen. For biochemical identification, according to ISO 6579 (2002) method: purified isolates were examined by different biochemical reaction based on oxidase, urea hydrolysis, H2s production on triple sugar iron, indole, methyl red, vogus-proskaur and citrate utilization tests. For biochemical identification, salmonella isolates were serotyped according to Kauffman (1974) method (7) for the determination of somatic (O) and flagellar (H) antigens using salmonella antiserum (DENKA SEIKEN Co., Japan).

Molecular detection of salmonella virulent genes by multiplex PCR

Bacterial DNA was extracted using QIAamp extraction kit following manufacturer's protocol and as previously described (8). Multiplex PCR was applied to amplify salmonella virulence genes *invA* and *hila* using specific primers as shown in table 1. PCR reaction mixture (25 µl) contained 5 µl of bacterial DNA, 2.5 µl of 10x Master mix (containing 1.25 U of Taq DNA polymerase, 1.5 mM MgCl2 and 2 µl 10mM dNTP mix), 1 µl of each primer (10 pmol) 15.5 µl DNase free water. The PCR cycling protocol was applied as following: an initial denaturation at 94°C for 60 sec, followed by 35 cycles of denaturation at 94°C for 60 sec, annealing at 64°C for 30 sec and extension at 72°C for 30 sec, followed by a final extension at 72°C for 7 min. Finally, 5 µl of each amplicon was electrophoresed in 1.5 % agarose gel, stained with ethidium bromide and visualized and captured on UV transilluminator. A 100 bp DNA ladder was used as a marker for PCR products.
Molecular detection of invA and hilA virulent genes in salmonella serovars isolated from fresh water fish

Results

The prevalence of salmonella species

The total prevalence rate of isolated salmonella was 32.5% (39/120) in all fish with highest incidence rate in Mugil species (47.5%, 19/39) followed by Clarias lazera (30%, 12/39) and Tilapia niloticus (20%, 8/30).

Serotyping

Salmonella isolates (n = 39) were serotyped using "O" and "H" antisera and the results showed 7 different salmonella serotypes: S. typhimurium, S. enteritidis, S. infantis, S. virchow, S. Heidelberg, S. wingrove, and S. tsevie with a total percentage of 33.3%, 23%, 12.8%, 12.8%, 10.25%, 5% and 2.5%, respectively. The prevalence of each serotype in each fish type was shown in Table 2.

Incidence of virulence genes

The incidence of invA and hilA virulent genes among the 7 salmonella serotypes as detected by multiplex PCR revealed prevalence of invA gene in all salmonella serotypes with a percentage of 100%, hilA gene in only 66.6% of all serotypes (Table 3 and Fig. 1).

Table 1: The primer sequence of salmonellae used for PCR

<table>
<thead>
<tr>
<th>Target gene</th>
<th>Oligonucleotide sequence (5’ → 3’)</th>
<th>Product size (bp)</th>
<th>References</th>
</tr>
</thead>
<tbody>
<tr>
<td>invA (F)</td>
<td>5’ GTGAAATTATCGCCACGTTCGGGCA ′3</td>
<td>284</td>
<td>(9)</td>
</tr>
<tr>
<td>invA (R)</td>
<td>5’ TCATGCACCGTCAAGGAACC ′3</td>
<td></td>
<td></td>
</tr>
<tr>
<td>hilA (F)</td>
<td>5’ CTGCCGCAGTGTTAAGGATA ′3</td>
<td>497</td>
<td>(10)</td>
</tr>
<tr>
<td>hilA (R)</td>
<td>5’ CTGTCCGCTTAATCGCATGT ′3</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

Table 2: Serotyping of salmonellae isolated from the examined samples of fresh water fishes

<table>
<thead>
<tr>
<th>Serotypes</th>
<th>Tilapia niloticus</th>
<th>Mugil cephalus</th>
<th>Clarias lazera</th>
<th>Group</th>
<th>Antigenic structure</th>
</tr>
</thead>
<tbody>
<tr>
<td>S. typhimurium</td>
<td>3(7.5)</td>
<td>6(15)</td>
<td>4(10)</td>
<td>B</td>
<td>1,4,5,12 i :1,2</td>
</tr>
<tr>
<td>S. infantis</td>
<td>1(2.5)</td>
<td>1(2.5)</td>
<td>3(7.5)</td>
<td>C1</td>
<td>6,7 r :1,5</td>
</tr>
<tr>
<td>S. virchow</td>
<td>2(5)</td>
<td>1(2.5)</td>
<td>2(5)</td>
<td>C1</td>
<td>6,7,14 r :1,2</td>
</tr>
<tr>
<td>S. enteritidis</td>
<td>2(5)</td>
<td>5(12.5)</td>
<td>2(5)</td>
<td>D1</td>
<td>1,9,12 g,m</td>
</tr>
<tr>
<td>S. heidelberg</td>
<td>0</td>
<td>3(7.5)</td>
<td>1(2.5)</td>
<td>B</td>
<td>1,4,5,12 r :1,2</td>
</tr>
<tr>
<td>S. tsevie</td>
<td>0</td>
<td>1(2.5)</td>
<td>0</td>
<td>B</td>
<td>4,5 i:e,n,z15</td>
</tr>
<tr>
<td>S. wingrove</td>
<td>0</td>
<td>2(5)</td>
<td>0</td>
<td>C2</td>
<td>8,20 c:1,2</td>
</tr>
<tr>
<td>Total (120)</td>
<td>8(20)</td>
<td>19 (47.5)</td>
<td>12(30)</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

Table 3: Incidence of virulence genes of different salmonella strains isolated from the examined samples of fish.

<table>
<thead>
<tr>
<th>Salmonella serovars</th>
<th>No. of isolates</th>
<th>invA No (%)</th>
<th>hila No (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>S. typhimurium</td>
<td>4</td>
<td>4 (100)</td>
<td>3 (75)</td>
</tr>
<tr>
<td>S. infantis</td>
<td>4</td>
<td>4 (100)</td>
<td>2 (50)</td>
</tr>
<tr>
<td>S. virchow</td>
<td>4</td>
<td>4 (100)</td>
<td>2 (50)</td>
</tr>
<tr>
<td>S. enteritidis</td>
<td>3</td>
<td>3 (100)</td>
<td>3 (100)</td>
</tr>
<tr>
<td>S. heidelberg</td>
<td>1</td>
<td>1 (100)</td>
<td>1(100)</td>
</tr>
<tr>
<td>S. tsevie</td>
<td>1</td>
<td>1 (100)</td>
<td>1(100)</td>
</tr>
<tr>
<td>S. wingrove</td>
<td>1</td>
<td>1 (100)</td>
<td>0</td>
</tr>
</tbody>
</table>
Figure 1: Agarose gel electrophoresis of multiplex PCR of \( invA \) (284 bp) and \( hilA \) (497 bp) virulence genes for characterization of salmonella species. Lane M: 100 bp ladder; lane C+: Control positive strain for \( invA \) and \( hilA \) genes; lane C-: Control negative; lanes 1-18 isolated samples.

Discussion

In the present study, we found high incidence of salmonella pathogen among different fish species with highest rate in \( Mugil cephalus \) (47.5%), followed by \( Clarias lazera \) (30%), and then \( Tilapia niloticus \) (20%). Higher incidence in \( Mugil cephalus \) may be due to its feeding nature as omnivorous bottom feeders swim over sandy-muddy bottom were they greases the pond detritus, organic matter and sediments which are usually of a high bacterial count (11) and salmonella can survive (54 days in water and up to 119 days in sediment) (12). This also may be due to the higher exposure of mugil species to contamination during their long transportation at high temperature from far cities as the point of production (farms) is so far from the point of distribution (fish markets) in Gharbia governate. On the other hand, tilapia mainly inhabited shallow streams which are usually of a low bacterial count as a result of higher speed of water flow which prevent exceeding contamination with microorganisms, indicating the possibility of occurring cross contamination during offloading, landing and marketing of tilapia fish besides applying of poor personal hygiene of fish handlers during sorting and selling, the degree of utensils contamination, the bacterial load of ice used for fish preservation and the time of sampling (16).

The results obtained from serological identification of the examined salmonella isolates revealed presence of 7 different salmonella serovars which are \( S. typhimurium \), \( S. enteritidis \), \( S. infantis \), \( S. virchow \), \( S. Heidelberg \), \( S. wingrove \) and \( S. tsevie \) with a percentage of 33.3%, 23%, 12.8%, 12.8%, 10.25%, 5% and 2.5%, respectively. These results agreed with (17) who reported that \( Salmonella typhimurium \) was the most frequently serovar isolated from.
Nile tilapia from Winam Gulf of lake victoria, kenia with a percentage of 14.3% regarding other salmonella serovars indicating its ability for survival and adaptation in the aquatic environment. This also was in consistence with WHO, (18) which reported that Salmonella typhimurium is the most common salmonella serotype isolated from cases of food poisoning and represents about 50-60% of such cases. Many of these incidents have been linked to consumption of raw or improperly cooked fish and fish products.

It has been found that salmonella has a large number of genes which implicated in its virulence. Many of these virulent genes are chromosomal genes located on the pathogenicity islands referred to as salmonella pathogenicity island (spl) which is a part of type 3 secretion systems (T3SS) and encoded genes like invA, and hilA allowing some salmonella species to invade the host cells and induce infection. Our obtained PCR results revealed that invA gene was present in all the isolates with a percentage of 100%. Meanwhile, hilA gene was present only in 66.6% of all the isolates. Other studies have reported similar results (19-22) indicating that invA gene can be used as a specific target for detection of salmonella as they are widely distributed among salmonella serovars reflecting the potential virulence of these isolates. These finding play a great role in determining the severity of the microbe since invA gene was found to be present and functional in most if not all salmonella serotypes responsible for the invasion of the cells of the intestinal epithelium allowing salmonella pathogen to enter and survive inside the eukaryotic cells with subsequent diseases in variety of hosts. In this regard, it has been used in determining the presence and metabolic activity of salmonella species (23). Meanwhile, hilA gene is required for regulation of type 3 secretion apparatus genes which secret proteins that are related to cell invasion [13]. And has been used in detecting of S.enterica serovars from fecal samples according to [19]. In recent studies, it has been used as a diagnostic target to validate alternative methods for food analysis and food-borne pathogen detection instead of the standard method of ISO 6579, 2004 which are time consuming and can take up to 4-6 days for confirmative results.

Conclusion

Presence of salmonella as enteropathogens in fresh water fish can reflects the poor hygienic conditions under which fish have been captured, handled and transported. The main finding in our study was the high prevalence of salmonella pathogen with a diversity of serotypes in fresh water fish in gharbia governorate creating a high level of attention towards the hazards of presence of these zoonotic pathogens in marketed fish and their bad impact on public health. so, our study recommends a monitoring and surveillance programs to be adopted against the food borne-salmonella by applying a strict hygienic measures under the supervision of the food controlling authorities in order to control the keeping quality of the locally consumed fish and to protect the consumer from the risk of food poisoning associated with consumption of contaminated fish with such dangerous pathogens.

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References


