

LAYING PERFORMANCE, FAT DIGESTIBILITY AND LIVER CONDITION OF LAYING HENS SUPPLEMENTED WITH VITAMIN B₁₂ OR BIOTIN AND/ OR BILE ACIDS IN DIET

Mohamed I. El-Katcha, Mosaad A. Soltan, Karima El-Naggar*, Set A. El-Shobokshy, Mohamed A. El-Erian

Department of Nutrition and Veterinary Clinical Nutrition, Faculty of Veterinary Medicine, Alexandria University, 22758, Egypt

*Corresponding author: karima.muhammad@alexu.edu.eg; ORCID ID: 0000-0002-7847-7613

Abstract: For 8 weeks feeding trial, two hundred and seventy, 53 weeks old laying hens were used to investigate the effects of dietary supplementation of vitamin B₁₂ or biotin and/or bile acids on performance, egg quality, fat digestibility and liver composition and histopathology. Birds were randomly divided into 6 groups (three replicates) and fed the experimental diets; group1 (G1) fed on the basal diet without additives as control while G2 and G3 supplemented with 0.02 ppm vitamin B₁₂ and 0.3 mg biotin/kg diet respectively, groups 4-6 fed as the previous detailed design of G1- G3 with the addition of 400 g of dried bile acid (DBA) /tone feed. Biotin supplementation significantly ($P<0.05$) increased body weight losses of laying hens, both vitamins significantly ($P<0.05$) decreased daily feed intake (FI) and improved FCR. DBA addition alone or with biotin reduced these weight losses along with significant ($P<0.05$) increase in daily FI. Vitamin B₁₂ supplementation alone or with DBA increased average egg production ($P<0.05$) while was reduced with biotin supplementation. Fat digestibility was non-significantly improved ($P\geq 0.05$) with both vitamins supplementation without or with DBA. Biotin significantly ($P<0.05$) reduced the average yolk relative weight, which was increased when mixed with DBA, while significantly increased average egg albumin relative weight. Vitamin B₁₂ or biotin addition without or with DBA non-significantly increased blood serum GOT and GPT activities, non-significantly reduced ($P\geq 0.05$) fat content of liver tissue (on dry matter or fresh basis) and serum lipid profile parameters except serum HDL concentration, was increased, with no histopathological changes in hepatic tissue. It could be summarized that vitamin B₁₂ supplementation without or with DBA is recommended in layer diet as it gave the best performance, reduced serum lipid profile and improved fat digestibility and the hepatic health.

Key words: layers; egg quality; liver; vitamin B₁₂; biotin; bile acids

Introduction

Recently, laying hens facing many challenges influencing egg production industry resulting in significant economic losses. Such

challenges as environmental changes, imbalanced or poor-quality feed, frequent diseases affect the physiological condition of layers and bring out pathological burden to hen body especially to sensitive and important organs such as liver. Therefore, liver is in continuous overloaded condition, which disturbs its function. One of the common disorders influences liver in layers is the fatty liver hemorrhagic syndrome (FLHS), negatively affecting their performance. FLHS is a metabolic or nutritional disorder of layers characterized by excessive accumulation of fat depots in the hepatic tissue and abdominal cavity as liver become enlarged, pale and ends up with rupture and hemorrhage causing death of the affected birds (1). This syndrome occurs when the lipogenesis process exceeds the capacity of lipoprotein synthesis and secretion (2). The actual cause of FLHS has not been fully defined, but could be a combination of nutritional, genetic, environmental, and hormonal factors (3). Different studies have been done to follow up the actual factors inducing this problem in laying hens with the development of some managerial and nutritional approaches to reduce its occurrence. One of these strategies is dietary supplementation of some feed additives such as some lipotropic factors and antioxidants, which could control the lipid metabolism, reduce the production of free radicals and protect the liver from damage.

Methionine and choline as lipotropic factors are used to treat or prevent this syndrome as their inadequate dietary levels increase oleic acid absorption and decreased triacylglycerol secretion (4, 5). Other nutrients deficiency such as vitamin B₁₂ and vitamin E increased hepatic triacylglycerol accumulation in laying hens (6, 7), increasing the incidence of occurrence. Earlier studies reported that biotin deficiency is an important dietary factor involved in inducing fatty liver in birds (8, 9).

Recently, the poultry production sector is continuously searching for new feed additives to improve feed efficiency and physiological condition of layers for optimal performance and egg production. Bile acids are natural components synthesized in human and animal body from cholesterol, which have a critical role in

dietary fat digestion and absorption through the intestinal wall (10). Previous studies have been done investigating impacts of B-vitamins supplementation and emulsifiers as bile acids on broilers health and productivity (11-14), however, little literature is available regarding their effect on laying hens productive performance and egg quality parameters.

We hypothesized that dietary inclusion of these additives could help in the digestion of fat, maintain the healthy condition of the liver of laying hens. Therefore, the main objectives of the present study to investigate the effects of vitamin B₁₂ or biotin supplementation without or with dried bile acids on productive performance, egg quality, fat digestibility, serum biochemistry and liver histopathology of laying hens.

Material and methods

Birds, experimental design and feeding program

Management procedures for all birds used during the course of experiment followed the guidelines approved by the Animal Care and Use Committee, Faculty of Veterinary Medicine, Alexandria University. A total of two hundred and seventy, 53 weeks old of *Isa brown* laying hens purchased from local company and used in this experiment. Hens were weighed separately, then randomly divided into 6 separate groups (45 bird /group) with three replicates each (15 bird/ replicate). Birds were kept in wire cages, and each cage was provided with feeder and water troughs. The control group (Group 1) was fed the basal diet (BD) without any additives, group 2 and 3 were fed the BD supplemented with 0.02 ppm vitamin B₁₂ and 0.3 mg biotin/kg diet respectively, while group 4-6 were fed as the previous detailed design of the first three groups but, with the addition of 400 g bile acid/tonne diet. BD were formulated to meet bird nutrient requirements according to NRC (15). BD composition and its chemical analysis is presented in table 1, which was analyzed according to AOAC (16). Additives used in this experiment included; vitamin B₁₂ and biotin produced by Allgäu Vet. Co., China; the dried bile acids (DBA) (Shangdong Longchang

Animal Health Product Co., Ltd, Jinan, China). All the experimental groups were fed the designated diets with free access to fresh water for 8 weeks experimental period. Hens were weighed individually at the beginning and end of experiment and the live body weight (BW) changes were taken with recording of feed intake (FI). Feed conversion ratio (FCR) was calculated based on feed consumed (g) and produced egg mass (g).

Egg production and Egg quality

Eggs produced from various laying hens were collected daily for calculating the average daily egg production % (EP) and egg weight (g) for each group for 14 days period for four successive periods. Egg mass was calculated by multiplying egg weight by egg production. At the end of the experiment, 20 eggs from each group were collected to estimate the egg quality parameters (yolk weight and relative weight to total egg weight, yolk index, albumin weight, and its relative weight, albumen index, shell thickness and weight as well as its relative weight were recorded) according to Card and Nesheim (17). Six samples of egg yolk from each group were collected for egg yolk cholesterol determination according to Rotenberg and Christensen (18).

Fat digestibility

During the last week of the feeding trail, the excreta were quantitatively collected for 5 successive days in addition to recording the amount of feed consumed. The excreta then dried in hot air oven, ground and stored until chemical analysis of fat. Ether extract of feed, fecal and liver samples was determined according to AOAC (16).

Serum biochemical parameters

Blood samples were collected (n= 6/group), then left to coagulate at room temperature. Separation of serum was done by centrifugation at 3000 rpm for 10 minutes. The clear serum samples were kept in freezer (-20°C) until analysis of the following blood serum indices; total cholesterol, high-density lipoprotein (HDL), low-density lipoprotein (LDL), triglyceride (TG), glutamic oxaloacetic transaminase (GOT) and

glutamic pyruvic transaminase (GPT). All these parameters were determined by spectrophotometer using specific commercial kits produced by Biodiagnostic, Co. (Diagnostic and Research reagents).

Liver Histopathology

At the end of the experiment (61 week of hen age), three hens from each group (one hen from each replicate) were collected randomly and used for liver tissue collection. Samples were fixed in 10% formalin for at least 2 days then slides were prepared according to Bancroft et al. (19), and stained with Hematoxylin and Eosin (H&E) for general inspection.

Statistical analysis

Results were analyzed using analysis of variance (one-way ANOVA) using IBM SPSS Statistics 22 statistical package (SPSS Inc., Chicago, IL, USA) to measure the significant differences between the means of different variables. Differences among experimental groups were considered significant at ($P<0.05$).

Results and discussion

Layer Performance

Biotin supplementation significantly ($P<0.05$) increased BW losses of laying hens. Both vitamins significantly ($P<0.05$) decreased average daily FI, but, improved FCR compared with control. On the other hand, DBA addition alone or with biotin reduced these losses in BW along with significant ($P<0.05$) increase in daily FI compared with laying hens without DBA addition (table 2). Moreover, FCR was significantly improved with DBA inclusion with vitamin B₁₂ and biotin compared with birds fed the same diet without DBA or control group.

Unlike the obtained results, earlier studies examined the effects of vitamin B₁₂ (20, 21), concluded that it had no clear effect on FI of commercial laying hens, while increased FI with Halle and Ebrahim (22). Improvement of FCR with vitamin B₁₂ supplementation in the present study is in line with Halle and Ebrahim (22), while inconsistent with Kato et al. (20). Additionally, Abdel-Mageed and Shabaan (23) documented another different result as similar

FI was obtained in laying hens groups fed biotin-supplemented diets while, improved FCR with increasing its level from 162.5 to 325.5 µg/kg. Additionally, Whitehead (24), found that biotin supplementation during lay did not have any beneficial effect upon FI and FCR of laying hens.

Bile acids are the principal constituents of bile, playing a significant role in emulsification, digestion, and absorption of fat and lipid-soluble vitamins (25). Best FCR was obtained with birds fed BD contained DBA plus vitamin B₁₂ (2.08), while the worst value with birds fed BD supplemented with DBA (2.49). Previous reports documented improvements of broiler performance (weight gain, higher FI and FCR) with the dietary inclusion of bile acids (13, 26-28), improved FCR in quail laying hens (29). The variation in response may be related to different breeds considered in these trials.

Regarding EP (table 2), vitamin B₁₂ supplementation alone or with DBA increased average EP ($P < 0.05$) with the highest EP (86.34%) in birds supplemented with the 2 additives suggesting that they could have a synergistic effect. On the other hand, biotin supplementation showed dissimilar result as it reduced EP (72.71%), even when added with DBA (78.22%), compared with control birds. Unlike the obtained result, Kato et al. (20) found that vitamin B₁₂ supplementation had no effect on EP, attributing it to short experimental period. Additionally, Bunchasak and Kachana (30) reported that supplemental vitamin B₁₂ didn't alter egg production or FI. However, Daryabari et al. (31) reported that 0.3g biotin/L water supplementation improved EP of breeding hens, as well as Abdel-Mageed and Shabaan (23) who found that biotin inclusion in diet enhanced it. Biotin concentration in the blood has been reported to be lower in young broiler breeder hens and increases linearly with age (32), therefore, the lower or unaffected EP in the biotin supplemented groups in the present study could be attributed to the higher level of biotin in birds. Response due to bile acids addition is inconsistent with the finding obtained with (29) as 1.2% supplemental ox bile addition in quail laying hens diet significantly improved EP %.

Both vitamins supplementation non-significantly ($P \geq 0.05$) reduced average egg weight (table 2). The same response was obtained when diet was supplemented with DBA alone or combined with vitamin B₁₂ while, when DBA mixed with biotin showed slightly higher egg weight compared with birds fed on the same diet without DBA. On contrary, Kato et al. (20) found increased egg weight of laying hens with vitamin B₁₂ supplementation.

Fat digestibility

Fat digestibility showed non-significant improvement ($P \geq 0.05$) with vitamin B₁₂ or biotin compared with control (table 2). The same result was obtained when DBA was supplemented alone or with both vitamins. Fat digestibility improvement especially in laying hens supplemented with DBA being the highest was supported by Alzawqari et al. (28), who found that supplemental increased levels of bile in diet linearly increased fat digestibility in broiler chicks. This improvement with the addition of DBA might be attributed to inadequate amount of bile salts secreted by bird, so more emulsification of fats occur by the exogenous bile acids or for replacement of the active catabolism of bile salts occur by gut microflora (26). Moreover, Lai et al. (12) indicated that supplementation with 60 and 80 mg/kg bile acids significantly increased the activity of intestinal lipase and lipoprotein lipase on day 21 and 42. The present study revealed a close relationship between dietary DBA supplementation and fat digestibility. Most of the previous studies reported, were concerned with investigation of the effects of these exogenous emulsifiers in broiler diet especially at younger age of chicks due to the physiological limitation of the digestive tract to produce endogenous emulsifiers. However, literature on their application in layers diets is scarce.

Egg quality (External and internal quality)

Average egg shell relative weight was non-significantly increased ($P \geq 0.05$) with vitamin B₁₂ or biotin compared with the reference group (table 3). DBA addition single or combined with vitamin B₁₂ increased ($P \geq 0.05$) egg shell

relative weight, while, was reduced ($P \geq 0.05$) when biotin was added to diet. Regarding egg shell thickness, no changes were found among different treatments. In support, Keshavarz (33) concluded that vitamin B₁₂ supplementation without or with methionine improved egg shell quality, Rajalekshmy (34) reported that shell weight was decreased at higher levels of folic acid (4 ppm) when vitamin B₁₂ was added (0.01 ppm). However, Kato et al. (20) found another different result as egg shell thickness and weight per unit of surface area were higher ($p < 0.01$) for birds fed diet without vitamin B₁₂ supplementation.

In terms of internal egg quality (table 3), biotin supplementation significantly reduced ($P < 0.05$) average egg yolk relative weight, but this effect was reversed when DBA was added with biotin. On the other hand, average egg albumin relative weight was significantly increased with biotin supplementation, though the lowest relative weight was recorded with birds received diet supplemented with DBA only. Regarding average yolk index, it was significantly reduced in birds supplemented with biotin and DBA, with no significant difference between the other experimental groups. The present data are in line with El-Husseiny et al. (35), who found that dietary supplementation with 0.01-0.02 ppm vitamin B₁₂ did not have any significant effect on egg shell thickness, shell percentage, Haugh units and yolk index. Moreover, Whitehead (24), reported that albumen height was slightly improved with biotin supplementation. Additionally, Abdel-Mageed and Shabaan (23), showed that biotin inclusion at 325.5 µg/kg diet gave the highest increase in egg shape index, yolk index and Haugh unit. Furthermore, Soltan (29) concluded that ox bile addition in quail laying hens had no significant influence on internal and external egg quality.

Egg yolk triglyceride and cholesterol content were non-significantly ($P \geq 0.05$) reduced with BDA supplementation compared with birds fed the same diets without DBA addition. These finding could support that DBA has an important role in fat metabolism through enhancing its digestibility and utilization. The egg is a highly nutritious food for human consumption,

however, its higher content of cholesterol associated with cardiovascular diseases restricts its consumption. So, previous studies were concerned with the production of egg with low cholesterol content through genetic selection; dietary interventions or using some drugs such as bile acids sequestrants in the diet of laying hens (36, 37).

Serum biochemical parameters

Blood serum lipid profile (table 4) was non-significantly altered with additives used. However, serum total cholesterol, TG, LDL, and VLDL concentrations were non-significantly ($P \geq 0.05$) reduced with DBA addition single or in combination with vitamins used except for serum HDL concentration which was increased. The higher reduction was observed in birds supplemented with DBA.

Serum TG, LDL, and HDL concentrations considered an important diagnostic indicators of lipid metabolism. The synthesis of adipose tissue, fat deposition, and formation of yolk in poultry is dependent on available serum TG. Most of fatty acids are produced in the liver and transported via LDL or chylomicrons for storage in adipose tissue as TG (2). On the other hand, HDL enhances the uptake and transport of cholesterol from extra hepatic tissues to hepatic tissue for catabolism (38), which was confirmed in the current study as it was non significantly increased especially with DBA addition. Therefore, these findings suggesting that the used feed additives (vitamin B₁₂, biotin and DBA) could have a hypolipidemic effect by affecting lipid metabolism. On contrary, Alzawqari et al. (13), reported that dietary inclusion of DBA significantly increased serum components of cholesterol, TG, HDL and LDL concentrations at days 21 and 42 of age, however, serum cholesterol concentration was numerically decreased (28). Furthermore, Lai et al. (12) documented that bile acid inclusion in broiler diets showed no differences in serum TG, HDL, and LDL concentrations among all groups at days 21 and 42. Inconsistency in results between experiments maybe related to differences in breed, age of birds and feeding experimental design.

Vitamin B12 or biotin supplementation without or with DBA non-significantly increased blood serum GOT and GPT activities compared with their reference birds (table 4). Serum GOT and GPT activity are traditional biochemical indices of liver function and are used clinically for diagnosis of liver injury. Yousefi et al. (5), documented that AST enzyme activity (GOT) could be used as diagnostic indicator for FLHS in laying hens. All liver function related parameters in the present study were within normal range (39), which could confirm that these supplemental additives in laying hen diet had no adverse effects on liver function.

Liver composition and histopathology

As presented in table 4, both vitamin supplementation without DBA addition non-significantly decreased liver moisture % while significantly increased ($P < 0.05$) it when combined with DBA compared with their reference birds. On the other hand, the fat content of the liver tissues (on dry matter or fresh basis), was non-significantly reduced ($P \geq 0.05$) with both vitamins supplementation without or with DBA. Fatty liver is associated with the reductions in egg production with poor egg shell quality (40), which is particularly a serious problem during the late period of laying. With the long-term accumulation of fat and various toxins in the hepatic tissue, hepatocytes are injured with

impaired function, reducing absorption and utilization of fat-soluble vitamins which consequently affecting the assimilation and deposition of calcium resulting in poor egg shell quality. So, liver health is closely associated with egg production and egg shell quality. Dietary inclusion of bile acids enhances the clearance function of very low-density lipoproteins, which transports fat from liver to systemic tissues, and reduces fat deposition in hepatocytes. Additionally, it stimulates the thyroid hormone activity (41), increases energy expenditure, reducing excessive accumulation of fat inside the liver and other body tissues of laying hens.

The examined liver of laying hens fed on the BD without any supplemental additives showed centrilobular vacuolation represented with clear vacuoles with round border consistent with fatty changes (Fig.1A). Most of the hepatic sinusoids were congested in addition to multifocal lymphocytic cells aggregation. Vitamin B₁₂ or biotin supplementation markedly decreased these pathological vacuolations with nearly normal hepatocytes (Fig.2 B and C). DBA inclusion in the diet without or with vitamin B₁₂ or biotin supplementation showed mild vacuolation of hepatocytes or even normal hepatocytes (Fig.1D, E, and F). The present findings are in line with Lai et al. (42), who concluded that liver of broiler chickens supplemented with bile acids was normal and no histological changes. Therefore, these findings revealed that these feed additives especially bile acids could help in maintaining liver health by reducing fat deposition in the hepatic tissue.

Table 1: Ingredients composition and chemical analysis of experimental diet

Ingredients	%
Yellow corn ground	56.83
Soybean meal (44% CP)	20.0
Corn gluten meal	5.0
Wheat bran	5.0
Vegetable oils (sunflower oil)	1.25
Ground lime stone ¹	9.80
Monocalcium phosphate ²	1.30
Common salt	0.25
Vitamin premix ³	0.15
Mineral premix ⁴	0.15
Lysine ⁵	0.02
Methionine ⁶	0.10
Choline ⁷	0.10
Mycotoxin adsorbent	0.05
Chemical analysis (%)	
Moisture	11.65
Crude protein	17.07
Ether extract	4.87
Ash	12.76
Crude fiber	4.43
NFE ⁸	49.22
Calcium	3.72
Phosphorus	0.63
ME (Kcal/kg) ⁹	2746.88

¹Lime stone contains 37 % calcium and locally produced. ²Mono calcium phosphate contains 21% phosphorus and 17 % calcium. ³Vitamin premix: Each 1.5 kg contains: Vit. A (12000000 IU), vit. D (2000000 IU), vit. E (10 g), vit. K3 (2 g), vit. B1 (1 g), vit. B2 (5 g), vit. B6 (1.5 g), vit. B12 (10 g), nicotinic acid (30 g), pantothenic acid (10 g), folic acid (1g), biotin (50 mg), produced by Archar Daniels Midland Co., LL., USA. ⁴Mineral Premix: Each 1kg contain, Manganese 100000 mg, Zinc 600000 mg, Iron 30000mg, Copper 10000 mg, Iodine 1000 mg, Selenium 200 mg, Cobalt 100 mg. ⁵Lysine: 98% lysine hydrochloride, Shandong Longue Co., China. ⁶DL-methionine, Evonik Co. (guaranteed analysis 99.5% DL- methionine). ⁷Choline: choline chloride 60 % with vegetable carrier (corn powder) produced by Shandong pharmaceutical Co., China. ⁸Nitrogen free extract was calculated by difference. ⁹ME were calculated according to NRC (1994).

Table 2: Effect of dietary vitamin B₁₂ or biotin supplementation without or with dried bile acids on performance and fat digestibility of laying hens

Parameters	Supplementation						
	Control	Without DBA ¹			With DBA		
		Vitamin B ₁₂	Biotin	Control	Vitamin B ₁₂	Biotin	
Initial body weight (g/hen)	1851.82 ±21.07 ^a	1826.55 ±19.19 ^a	1815.66 ±23.77 ^a	1798.47 ±22.20 ^a	1844.27 ±19.63 ^a	1829.60 ±24.50 ^a	
Final body weight (g/hen)	1827.74 ±35.02 ^a	1802.24 ±29.49 ^{ab}	1731.11 ±31.14 ^b	1794.15 ±30.66 ^{ab}	1815.27 ±28.93 ^{ab}	1795.38 ±26.79 ^{ab}	
Weight changes (g/hen)	-24.08 ±20.02 ^b	-24.31 ±16.06 ^b	-84.55 ±19.26 ^a	-4.32 ±17.40 ^c	-29.00 ±17.35 ^b	-34.22 ±10.38 ^b	
Average Feed Intake (g/hen/day)	113.41 ±0.07 ^b	109.99 ±0.19 ^d	96.90 ±0.13 ^f	120.21 ±0.06 ^a	111.35 ±0.12 ^c	107.34 ±0.16 ^e	
Average FCR	2.35 ±0.03 ^b	2.14 ±0.03 ^c	2.14 ±0.06 ^c	2.49 ±0.03 ^a	2.10 ±0.03 ^d	2.19 ±0.03 ^c	
Average egg production (%) 53 – 61	76.65 ±0.69 ^{bc}	81.56 ±0.74 ^{ab}	72.17 ±1.37 ^c	78.60 ±0.79 ^b	86.34 ±4.27 ^a	78.22 ±0.76 ^b	
Average egg wt. (g)	62.83 ±0.22 ^a	62.74 ±0.15 ^a	62.48 ±0.15 ^a	61.38 ±0.16 ^a	61.75 ±0.14 ^b	62.65 ±0.14 ^a	
Average egg mass (g/hen)	48.17 ±0.46 ^{bc}	51.21 ±0.50 ^{ab}	45.20 ±0.89 ^c	48.21 ±0.48 ^{bc}	53.33 ±2.63 ^a	49.02 ±0.50 ^b	
Fat digestibility %	75.07 ±2.84 ^a	81.67 ±2.40 ^a	78.97 ±4.65 ^a	84.40 ±0.60 ^a	82.30 ±1.95 ^a	79.20 ±6.45 ^a	

Values are means ± standard error. Means within the same row of different litters are significantly different at ($P < 0.05$).¹ DBA: Dried bile acids

Table 3: Effect of dietary vitamin B₁₂ or biotin supplementation without or with dried bile acids on egg quality parameters

Parameters	Supplementation						
	Control	Without DBA ¹			With DBA		
		Vitamin B ₁₂	Biotin	Control	Vitamin B ₁₂	Biotin	
External egg quality							
Average eggshell relative weight	12.86 ±0.49 ^a	13.25 ±0.20 ^a	12.90 ±0.39 ^a	13.43 ±0.34 ^a	12.97 ±0.13 ^a	12.23 ±0.29 ^a	
Average eggshell thickness (mm)	38.78 ±0.36 ^a	37.56 ±0.47 ^a	37.04 ±0.70 ^a	37.19 ±0.64 ^a	37.94 ±1.42 ^a	37.33 ±0.94 ^a	
Internal egg quality							
Average yolk relative wt.	29.99 ±0.89 ^{ab}	29.82 ±0.50 ^{ab}	29.22 ±0.66 ^b	30.06 ±0.58 ^{ab}	30.74 ±0.69 ^{ab}	31.47 ±0.76 ^a	
Average yolk index	0.43 ±0.01 ^a	0.41 ±0.01 ^{ab}	0.43 ±0.00 ^a	0.43 ±0.00 ^a	0.41 ±0.01 ^{ab}	0.40 ±0.01 ^b	
Average albumin relative wt.	57.15 ±0.60 ^{ab}	57.17 ±0.39 ^{ab}	58.24 ±0.64 ^a	54.84 ±2.02 ^b	56.51 ±0.50 ^{ab}	55.17 ±0.82 ^{ab}	
Average albumin index	8.89 ±0.24 ^a	8.71 ±0.09 ^a	8.78 ±0.23 ^a	8.68 ±0.20 ^a	8.81 ±0.16 ^a	8.84 ±0.15 ^a	
Yolk cholesterol							
(mg/g yolk)	9.27 ±0.41 ^a	8.93 ±0.89 ^a	10.11 ±1.44 ^a	8.85 ±0.25 ^a	8.85 ±0.33 ^a	8.89 ±0.34 ^a	
(mg/whole yolk)	181.48 ±3.65 ^a	165.58 ±20.61 ^a	181.54 ±35.65 ^a	170.35 ±6.36 ^a	161.63 ±11.76 ^a	162.76 ±6.47 ^a	
Yolk triglycerides							
(mg/g yolk)	10.11 ±0.49 ^a	10.31 ±1.55 ^a	9.74 ±0.13 ^a	9.75 ±0.52 ^a	9.95 ±0.66 ^a	9.56 ±0.48 ^a	
(mg/whole yolk)	197.86 ±3.73 ^a	189.29 ±23.47 ^a	176.66 ±11.79 ^a	187.14 ±4.53 ^a	181.00 ±11.40 ^a	174.69 ±5.72 ^a	

Values are means ± standard error. Means within the same row of different litters are significantly different at ($P < 0.05$).¹ DBA: Dried bile acids

Table 4: Effect of dietary vitamin B₁₂ or biotin supplementation without or with dried bile acids on blood serum parameters of laying hens

	Supplementation					
	Control	Vitamin B ₁₂	Biotin	Control	Vitamin B ₁₂	Biotin
	Without DBA ⁶			With DBA		
Blood serum lipid profile parameters						
Cholesterol (mg/dl)	198.15 ±0.70 ^a	191.23 ±6.12 ^a	196.30 ±2.37 ^a	195.53 ±3.43 ^a	190.69 ±2.31 ^a	190.77 ±2.75 ^a
Triglyceride (mg/dl)	205.14 ±1.14 ^a	205.86 ±3.62 ^a	206.91 ±1.70 ^a	203.61 ±3.06 ^a	201.08 ±2.29 ^a	204.90 ±0.58 ^a
HDL ¹ (mg/dl)	47.17 ±2.06 ^a	48.43 ±2.10 ^a	47.23 ±1.57 ^a	49.80 ±1.42 ^a	50.80 ±2.60 ^a	51.63 ±1.49 ^a
LDL ² (mg/dl)	109.95 ±1.13 ^a	101.62 ±5.46 ^a	107.69 ±3.61 ^a	105.01 ±3.24 ^a	99.67 ±0.75 ^a	98.15 ±3.06 ^a
VLDL ³ (mg/dl)	41.03 ±0.23 ^a	41.17 ±0.72 ^a	41.38 ±0.34 ^a	40.72 ±0.61 ^a	40.22 ±0.46 ^a	40.98 ±0.12 ^a
Blood serum parameters related to liver functions						
GOT ⁴ (u/L)	65.33 ±17.9 ^a	96.33 ±3.18 ^a	71.33 ±16.51 ^a	94.00 ±2.08 ^a	82.50 ±12.50 ^a	77.00 ±10.50 ^a
GPT ⁵ (u/L)	20.90 ±3.51 ^a	24.67 ±1.45 ^a	21.50 ±0.50 ^a	21.67 ±1.45 ^a	22.50 ±0.50 ^a	24.67 ±1.67 ^a
Liver tissue composition (%)						
Moisture	70.37 ±0.53 ^{ab}	70.23 ±0.43 ^{ab}	69.99 ±2.54 ^{ab}	69.08 ±1.59 ^b	73.03 ±0.71 ^a	71.13 ±0.93 ^{ab}
Fat (DM basis)	25.43 ±3.30 ^a	22.78 ±2.47 ^a	19.62 ±3.50 ^a	22.86 ±4.18 ^a	21.51 ±1.67 ^a	23.61 ±0.99 ^a
Fat (fresh basis)	7.55 ±1.05 ^a	6.08 ±0.81 ^a	5.96 ±1.27 ^a	7.06 ±1.72 ^a	5.82 ±0.59 ^a	6.80 ±0.12 ^a

Values are means ± standard error. Means within the same row of different litters are significantly different at (P<0.05). ¹HDL: high-density lipoprotein, ²LDL: low-density lipoprotein (LDL), ³VLDL: very low-density lipoprotein, ⁴ GOT: glutamic oxaloacetic transaminase, ⁵GPT: glutamic pyruvic transaminase. ⁶ DBA: Dried bile acids

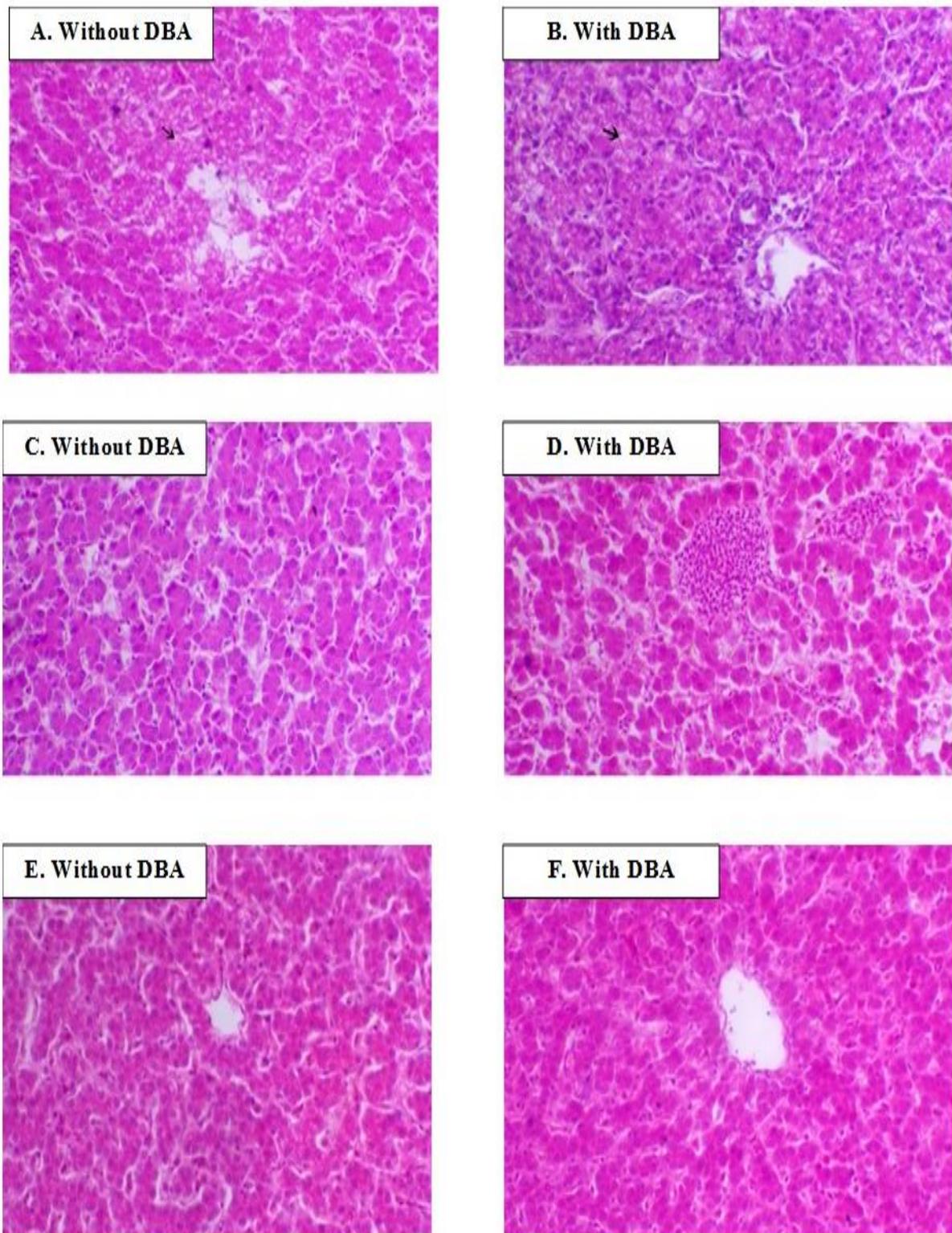


Figure 1: Histopathological changes of hepatic tissue of laying hens fed on control (without DBA) showing centrilobular hepatic vacuolation with fatty changes (arrow) (A); or with DBA mild degree of hepatic vacuolation consistent with fatty changes (arrow) (B); hepatic tissues of birds supplemented with vitamin B₁₂ supplementation and without DBA or with DBA addition showing normal hepatocytes (C and D); Birds supplemented with biotin without or with DBA addition showing normal hepatocytes (E and F)

Conclusion

Under the conditions of the present study, vitamin B₁₂ supplementation single or combined with dried bile acids is recommended in laying hen diet as it gave the best productive performance and improved fat digestibility, hepatic health and reduced the serum lipid profile. On the other hand, biotin supplementation is not recommended for laying hens except if combined with DBA.

Conflict of interest

All authors declared that no conflicts of interest, as they are responsible for the content and writing this article.

References

1. Crespo R, Shivaprasad HL. Developmental, metabolic, and other noninfectious disorders, in: YME Saif (Ed.), *Diseases of Poultry*, Iowa State Press, Ames, Iowa, 2003, 1055–102.
2. Hermier D. Lipoprotein metabolism and fattening in poultry. *J Nutr* 1997; 127(5 Suppl): 805s–8s.
3. Hansen RJ, Walzem RL. Avian fatty liver hemorrhagic syndrome: a comparative review. *Adv Vet Sci Comp Med* 1993; 37: 451–68.
4. Rinella ME, Elias MS, Smolak RR, Fu T, Borensztajn J, Green RM. Mechanisms of hepatic steatosis in mice fed a lipogenic methionine choline-deficient diet. *J Lipid Res* 2008; 49(5): 1068–76.
5. Yousefi M, Shivazad M, Sohrabi-Haghdoost I. Effect of Dietary Factors on Induction of Fatty Liver-Hemorrhagic Syndrome and its Diagnosis Methods with Use of Serum and Liver Parameters in Laying Hens. *Int J Poult Sci* 2005; 4: 568–72.
6. Couch J. Fatty livers in laying hens - a condition which may occur as a result of increased strain. *Feedstuffs* 1956: 28–46.
7. Leeson S. Metabolic Challenges: Past, Present, and Future. *J Appl Poult Res* 2007; 16(1): 121–5.
8. Whitehead CC, Blair R, Bannister DW, Evans AJ, Jones RM. The involvement of biotin in preventing the fatty liver and kidney syndrome in chicks. *Res Vet Sci* 1976; 20(2): 180–4.
9. Whitehead CC, Bannister DW, Evans AJ, Siller WG, Wight PAL. Biotin deficiency and fatty liver and kidney syndrome in chicks given purified diets containing different fat and protein levels. *Br J Nutr* 1976; 35(1): 115–25.
10. Reshetnyak VI. Physiological and molecular biochemical mechanisms of bile formation. *World J Gastroenterol* 2013; 19(42): 7341–60.
11. Quarantelli A, Bonomi A, Righi F, et al. The effects of different levels of dietary biotin on the performances and on bone growth in the broiler. *Ital J Anim Sci* 2003; 2 (sup1): 453–5.
12. Lai W, Huang W, Dong B, et al. Effects of dietary supplemental bile acids on performance, carcass characteristics, serum lipid metabolites and intestinal enzyme activities of broiler chickens. *Poult sci* 2018; 97(1): 196–202.
13. Alzawqari MH, Al-Baadani HH, Alhidary IB, Al-Owaimer AN, Abudabos AM. Effect of taurine and bile acid supplementation and their interaction on performance, serum components, ileal viscosity and carcass characteristics of broiler chickens. *S Afr J Anim Sci* 2016; 46: 448–57.
14. Cengiz Ö, Hess JB, Bilgili SF. Dietary biotin supplementation does not alleviate the development of footpad dermatitis in broiler chickens. *J Appl Poult Res* 2012; 21(4): 764–9.
15. NRC, *Nutrient requirements of poultry*. 9th Ed. Washington, DC: National Academic Press, 1994.
16. AOAC, *Official methods of analysis*. (18th ed.) Washington, DC: AOAC: Association of Official Analytical Chemists, 2005.
17. Card LE, Nesheim MC, *Poultry Production*, 11th Edn. Philadelphia: Lea and febiger Press, 1972.
18. Rotenberg S, Christensen K. Spectrophotometric Determination of Total and Free Cholesterol in Egg Yolk and Animal Tissues. *Acta Agric Sc and A* 1976; 26(2): 94–8.
19. Bancroft JD, Layton C, Suvarna SK, *Bancroft's Theory and Practice of Histological Techniques*. 7th edition. Churchill Livingstone: Elsevier, 2013.
20. Kato R, Bertechini A, Fassani E, Santos C, Dionizio M, Fialho E. Cobalt and vitamin B₁₂ in diets for commercial laying hens on the second cycle of production. *Braz J Poult Sci* 2003; 5: 45–50.
21. Squires MW, Naber EC. Vitamin profiles of eggs as indicators of nutritional status in the laying hen: vitamin B₁₂ study. *Poult sci* 1992; 71(12): 2075–82.
22. Halle I, Ebrahim M. Influence of Vitamin B₁₂ and Cobalt on performance of laying hens *Landbauforschung* 2012; 3(62): 111–6.

23. Abdel-Mageed MAA, Shabaan SA. Effect of supplemental biotin on the performance of aged fayoumi hens and progeny performance Egypt Poult Sci 2012; 32: 895–8.
24. Whitehead CC. Biotin- New information on requirements and supplementation in poultry, In Proceedings of Roche symposium, London, 1980.
25. Russell DW, Setchell KDR. Bile acid biosynthesis. Biochemistry 1992; 31(20): 4737–49.
26. Maisonnier S, Gomez J, Bree A, Berri C, Baeza E, Carre B. Effects of microflora status, dietary bile salts and guar gum on lipid digestibility, intestinal bile salts, and histomorphology in broiler chickens. Poult sci 2003; 82(5): 805–14.
27. Parsaie S, Shariatmadari F, Zamiri MJ, Khajeh K. Influence of wheat-based diets supplemented with xylanase, bile acid and antibiotics on performance, digestive tract measurements and gut morphology of broilers compared with a maize-based diet. Br Poult Sci 2007; 48(5): 594–600.
28. Alzawqari M, Moghaddam HN, Kermanshahi H, Raji AR. The effect of desiccated ox bile supplementation on performance, fat digestibility, gut morphology and blood chemistry of broiler chickens fed tallow diets. J Appl Anim Res 2011; 39(2): 169–74.
29. Soltan MA. Effect of diet containing *Nigella Sativa* (Black seeds) and/or ox bile on growth and productive performance of Japanese quail. AJVS 1999; 15(3): 655–69.
30. Bunchasak C, Kachana S. Dietary folate and vitamin B12 supplementation and consequent vitamin deposition in chicken eggs. Trop Anim Health Prod 2009; 41(7): 1583.
31. Daryabari H, Akhlaghi A, Zamiri MJ, et al. Reproductive performance and oviductal expression of avidin and avidin-related protein-2 in young and old broiler breeder hens orally exposed to supplementary biotin. Poult sci 2014; 93(9): 2289–95.
32. Whitehead CC. Biotin intake and transfer to the egg and chick in broiler breeder hens housed on litter or in cages. Br Poult Sci 1984; 25(2): 287–92.
33. Keshavarz K. Effects of reducing dietary protein, methionine, choline, folic acid, and vitamin B12 during the late stages of the egg production cycle on performance and egg shell quality. Poult sci 2003; 82(9): 1407–14.
34. Rajalekshmy PK. Effects of dietary choline, folic acid and vitamin B12 on laying hen performance, egg components and egg phospholipid composition (Ph.D.). University of Nebraska, Lincoln, 2010.
35. El-Husseiny OM, Soliman AZ, Omara II, El-Sherif HMR. Evaluation of Dietary Methionine, Folic Acid and Cyanocobalamin (B12) and Their Interactions in Laying Hen Performance. Int J Poult Sci 2008; 7: 461–9.
36. Mori AV, Mendonca JR, Watanabe C. Effects of cholestyramine and lovastatin upon plasma lipids and egg yolk cholesterol levels of laying hens. Braz J Vet Res Anim Sci 2000; 37: 00–00.
37. Ismail IB, Al-Busadah KA, El-Bahr SM. Effect of Dietary Supplementation of Canola Oil on Egg Production, Quality and Biochemistry of Egg Yolk and Plasma of Laying Hen. Int J Biol Chem 2013; 7: 27–37.
38. Miller NE, Nestel PJ, Clifton-Bligh P. Relationships between plasma lipoprotein cholesterol concentrations and the pool size and metabolism of cholesterol in man. Atherosclerosis 1976; 23(3): 535–47.
39. Marjanovic D, Kozic L, Petrovic M, Palic T, Litricin V. Serum transaminase activity in fowls. Vet Bull 1975; 45: 2890–6.
40. Trott KA, Giannitti F, Rimoldi G, et al. Fatty liver hemorrhagic syndrome in the backyard chicken: a retrospective histopathologic case series. Vet Pathol 2014; 51(4): 787–95.
41. Watanabe M, Houten SM, Mataka C, et al. Bile acids induce energy expenditure by promoting intracellular thyroid hormone activation. Nature 2006; 439(7075): 484–9.
42. Lai W, Cao A, Li J, Zhang W, Zhang L. Effect of High Dose of Bile Acids Supplementation in Broiler Feed on Growth Performance, Clinical Blood Metabolites, and Organ Development. J Appl Poult Res 2018; 27(4): 532–9.